Amino Acid Sequence Analysis of the Two Major Outer Capsid Proteins (VP7 and VP4) from Human-Derived Canine G3P[3] Rotavirus Strain Detected in Brazil

Análise da Seguência de Aminoácidos das Duas Principais Proteínas do Capsídeo Externo (VP7 e VP4) de Cepa de Rotavírus Humano Derivado de Cão G3P[3] Detectada no Brasil

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Abstract

Introduction: A close look at the rotavirus group A (RVA) genotypes in Brazil revealed the detection of a rare G3P[3] strain close related to canine strains. The aim of this study was to add to the already known genetic analysis by the description of the G3P[3] (IAL-R2638 strain) amino acid characteristics. Methods: Amino acid sequence analysis and protein based trees were conducted using BioEdit and MEGA 4.0. Results: The VP7 and VP4 protein of the IAL-R2638 strain displayed the highest amino acid identity to the canine-derived human strain HCR3A (99.2%), and to the canine strain RV52/96 (96.4%), respectively. IAL-R2638 strain did not possess an extra VP7 N-linked glycosylation site at amino acid 238 recently described for some G3 strains, as well as RotaTeqTM G3 vaccine strain. The topology exhibited by phylogenetic trees in previous analysis were maintained in the present amino acid-based trees, reinforcing a stable relationship between G3P[3] strains. Conclusions: Amino acid analysis data were consistent with the previous sequence of data obtained for the IAL-R2638 strain, supporting its possible canine origin. Theoretically, RotaTegTM vaccine could efficiently protect against G3P[3] infections based on the lack of the extra VP7 N-linked glycosylation site at amino acid 238. Phylogenetic analysis hypothesizes that all features undergo evolution independently of each other; however, unfavorable effects of nucleotide substitutions may be compensated by substitutions in other positions. The present study raises the question as to whether the amino acid-based trees could be applied as an approach to the study of RVA evolution, avoiding incorrect phylogenetic reconstructions.

Keywords: Disease Vectors. Epidemiological Surveillance. Gastroenteritis. Diarrhea. Rotavirus. Reoviridae Infections.

Resumo

Introdução: A vigilância epidemiológica dos rotavírus do grupo A (RVA) no Brasil possibilitou a detecção de cepa rara G3P[3] geneticamente semelhante às cepas caninas. O objetivo do presente estudo foi realizar a análise de aminoácidos da cepa G3P[3] (IAL-R2638) e complementar as informações adquiridas por meio da análise nucleotídica. Métodos: A análise da sequência de aminoácidos e a construção das árvores proteicas foi realizada utilizando os programas BioEdit e MEGA 4.0. Resultados: As proteínas VP7 e VP4 da cepa IAL-R2638 exibiram maior identidade de aminoácidos com a cepa humana derivada de cão HCR3A (99,2%) e com a cepa canina RV52/96 (96,4%), respectivamente. A cepa IAL-R2638 não apresenta o sítio adicional de glicosilação N na posição 238 na proteína VP7, recentemente descrito em algumas cepas G3, assim como a cepa G3 vacinal da vacina RotaTeqTM. As topologias exibidas nas árvores filogenéticas previamente analisadas foram mantidas nas árvores baseadas em aminoácidos, reforçando o parentesco estável entre cepas G3P[3]. Conclusões: A análise proteica da cepa IAL-R2638 foi consistente com os dados genéticos obitidos previamente, corroborando sua possível origem canina. Teoricamente, a vacina RotaTeq™ pode ser eficaz contra infecções por G3P[3], uma vez que essas cepas não possuem o sítio extra de glicosilação N 238. Análise filogenética pressupõe que todas as substituições nucleotídicas acarretam evolução de forma independente. Entretanto, substituições desfavoráveis podem ser compensadas por novas substituições em outras posições. O presente trabalho sugere o uso de árvores baseadas em aminoácidos como uma ferramenta auxiliar no estudo da evolução dos RVA, evitando construções filogenéticas incorretas.

Palavras-chave: Vetores de doenças. Vigilância epidemiológica. Gastroenterite. Diarreia. Rotavirus. Infecções por Reoviridae.

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INTRODUCTION

Group A rotavirus (RVA) are an important cause of severe gastroenteritis among infants and young children worldwide, as well as in animals of a wide variety of species.¹ RVA is classified into different P and G genotypes based upon the main neutralization antigens, namely, the spike protein (VP4) and the major outer capsid glycoprotein (VP7). The most prevalent genotypes, G1P[8], G2P[4], G3P[8], G4P[8], and G9P[8], are responsible for approximately 90% of worldwide human RVA infections.² However, RVA epidemiology is constantly changing, mainly due to reassortment events, and interspecies transmission of animal RVA to humans.³

With regard to the high incidence of RVA infections worldwide, two oral live-attenuated vaccines have been licensed. RotarixTM is a monovalent vaccine derived from a human G1P[8] strain, and has been introduced in Brazilian Immunization Program since 2006.^{4,6} RotateqTM is a human-bovine reassortant RVA vaccine that contains the human genotypes G1, G2, G3, G4, and P[8],⁶ and is available at private clinics. The post-marketing surveillance of circulating RVA genotypes is crucial for vaccine efficacy studies. In addition, selective vaccine pressure could also increase the circulation of uncommon animal-like strains.⁷ In fact, a close look at RVA genotypes in Brazil has already revealed the detection of uncommon G3P[3] strain with similarity to canine strains in 2012.⁸

Sequence similarities have been used as evidence for evolutionary relationships between strains. However, phylogenetic trees based on the analyses of DNA sequence may be misleading, especially when G+C content differs widely among lineages; therefore protein-based trees from amino acid sequences may be more reliable.^{9,10,11} Moreover; simulations on model proteins have shown that a smaller number of correlated substitutions is enough to change topology of the resulting phylogenetic trees.¹² The aim of this study was to add to the already known genetic analysis by the description of the G3P[3] (IAL-R2638 strain) amino acid characteristics.

MATERIAL AND METHODS

Strain

RVA IAL-R2638 strain (G3P[3]) was isolated from a 1 year old child with gastroenteritis symptoms in São Paulo, Brazil in 2011.⁸ Partial VP7 and VP4 sequences suggested a common origin between this human strain and canine strains.⁸ The amino acid characteristic of IAL-R2638 strain was determined in this study.

RNA extraction and reverse transcription-PCR (RT-PCR)

Viral RNA was extracted using QIAamp[®] Viral RNA Mini kit (Qiagen, Inc., Valencia, CA) according to the manufacturer's instructions. RT-PCR for VP7 and VP4 genes was performed according to the protocol previously described.^{13,14}

Nucleotide sequencing

The G3P[3] (IAL-R2638 strain) PCR amplicons were sequenced using the BigDye[™] kit v3.1 (Applied Biosystems, Inc., Foster City, CA) with primers Beg9 and End9 for the VP7 gene; Con2 and Con3 for the VP4 gene. Dye-labeled products were sequenced using an ABI 3130 sequencer (Applied Biosystems, Inc., Foster city, CA, USA).

Protein sequence analysis

The sequences obtained for the VP7 and VP4 (accession number JN848803 and JN848804, respectively) genes, and a set of cognate sequences of human and animal RVA available in the GenBank database were assembled with the BioEdit Sequence Alignment Editor (version 7.0.5.2) program. Protein sequence analysis was conducted using BioEdit and MEGA software version 4.0.¹⁵ Amino acid trees were constructed using the neighbor-joining method based on Poisson correction model.

Results

The deduced amino acid sequence of the VP7 gene from human strain IAL-R2638 was determined and compared to those of reference RVA strains belonging to the G3 genotype (Figure 1). The VP7 amino acid sequence of strain IAL-R2638 was 90.5 to 99.2% identical to those of RVA strains exhibiting G3 genotype specificity, and the highest amino acid identity (99.2% and 98.8%) was found to human RVA strain HCR3A (USA) and Ro1845 (Israel), respectively. Strain IAL-R2638 also exhibited high amino acid identity to canine strains A79-10 (98.8%), CU-1 (98.8%), RV198-95 (98.8%), K9 (98.4%), and to feline strain Cat97 (98.8%) (Figure 2). The lowest amino acid identity was observed between strain IAL-R2638 and murine strain MelMuRV (90.5%) (Figure 2).

The VP7 protein of strain IAL-R2638 had a potential N-linked glycosylation site located at amino acid 69 (Asn) (Figure 1). The antigenic regions A-F of strain IAL-R2638 clearly support its classification as genotype G3. Within the VP7 hyper variable regions A-F, the VP7 of human strain IAL-R2638 was completely identical to the canine strains A79-10, CU-1, and RV198-95; to the human strains HCR3A and Ro1845; and to the feline strain Cat97 (Figure 1). VP7 hyper-variable region B (amino acids 87 to 101) seems to be the most conserved region considering all strains analyzed, while region A (amino acid 39 to 50) seems to be the most variable (Figure 1).

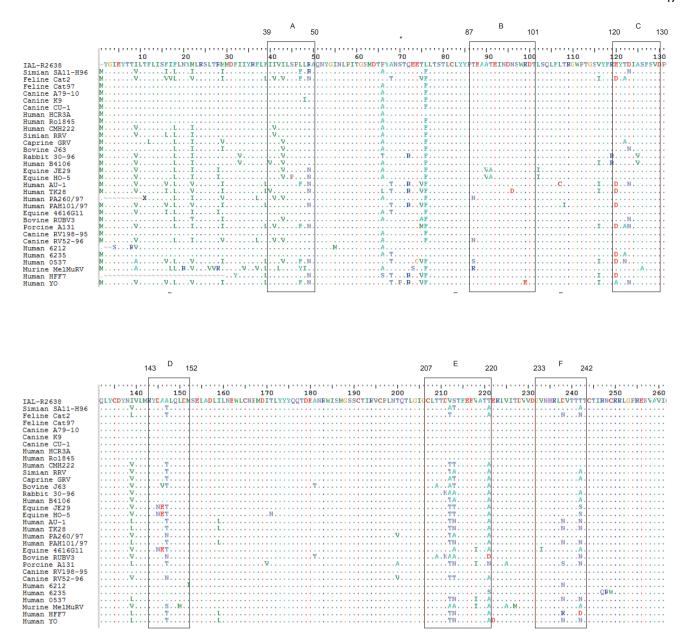


Figure 1 Deduced amino acid sequence of the VP7 protein of human strain IAL-R2638, and of a selection of G3 rotaviruses. The VP7 antigenic regions A-F are indicated. The glycosylation site NST (amino acids 69 to 71) is indicated by asterisks. Species and isolate of each strain are indicated.

The alignment of the deduced amino acid sequences from VP7 gene revealed amino acid substitutions inside the variable region D (amino acids 143 to 152) at position $147^{A \rightarrow T}$; region E (amino acids 207 to 220) at positions $212^{A/T \rightarrow V}$ and $213^{A/T/N \rightarrow V}$; and region F (amino acids 233 to 242) at position $242^{A/N/S \rightarrow V}$. Amino acid substitutions were also observed outside VP7 hyper-variable regions in IAL-R2638 strain: $66^{A \rightarrow P}$, $76^{F \rightarrow L}$, $221^{A \rightarrow T}$ and $278^{A \rightarrow V}$ (Figure 1). Among RVA strains with G3 genotype specificity, two distinct VP7 branches or groups were observed in amino acid-based tree, and designated A and B (Figure 2). The Brazilian RVA strain IAL-R2638 clustered into group A, along with human and animal RVA G3 genotypes from different countries, including the reference simian strains RRV (EU636932) and SA11-H96 (DQ838620) both from USA (Figure 2). Group B comprised most of the human G3 strains, excluding the porcine strain A131 (L35055) and the feline strain Cat2 (EU708961) (Figure 2).

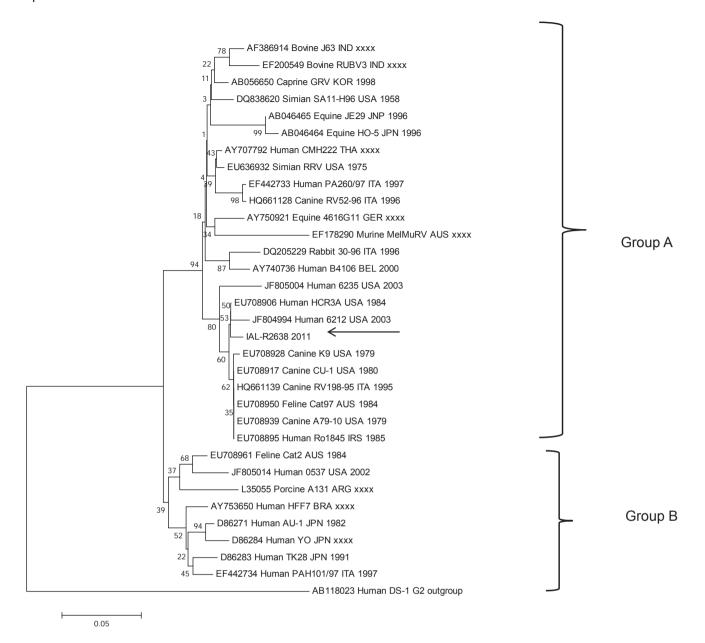


Figure 2 Neighbor-joining tree of the partial VP7 deduced amino acid sequence generate with MEGA 4.0 software of the IAL-R2638 strain (arrow). Reference G3 strains were obtained from GenBank database. Accession number, species, isolates, countries and year of each strain are indicated. Group A and B represent two clusters genetically distinct. The scale indicates the number of divergent amino acid residues. Percentages of bootstrap values are shown at the branch node.

Figure 3 shows the deduced amino acid sequence of the VP4 (subunit VP8*) of the human strain IAL-R2638 and representative VP4 amino acid sequences of RVA P[3] genotype. The potential cleavage sites, arginine (R) at positions 231 and 241, were maintained in strain IAL-R2638. The third arginine at position 247 in strain IAL-R2638 was substituted by a lysine (K). The same substitution was also observed in feline strains Cat97 and FRV64; in canine strains A79-10, K9, CU-1, and RV52/96; and in human strains HCR3A, Ro1845, 6212, and 6235. The highly conserved cysteines (C) at residues 203 and 216, and proline (P) at residues 68, 71, 225 and 226 were maintained in strain IAL-R2638 (Figure 3).

Comparative analysis of the deduced amino acid sequences of the strain IAL-R2638 VP4 fragment (VP8* subunit) showed that the variable region between amino acid 71 and 204 was fairly conserved among all strains analyzed, confirming the classification of strain IAL-R2638 as P[3] genotype. Within the VP8* subunit variable region, a unique and remarkable substitution occurred in strain IAL-R2638 at positions $148^{Q \rightarrow L}$ and $149^{N \rightarrow S}$ (Figure 3).

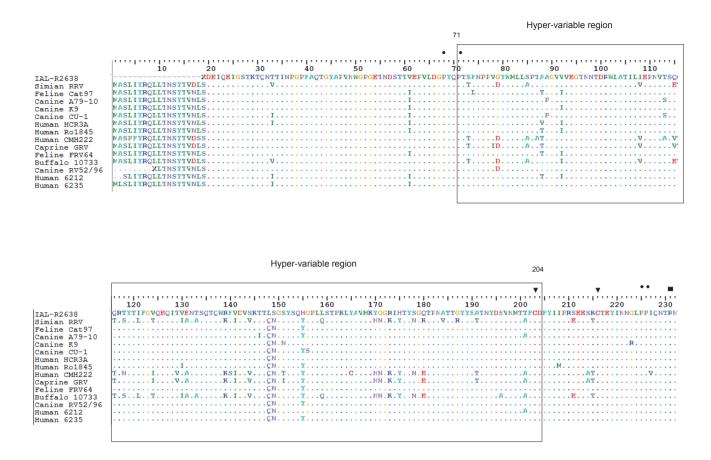
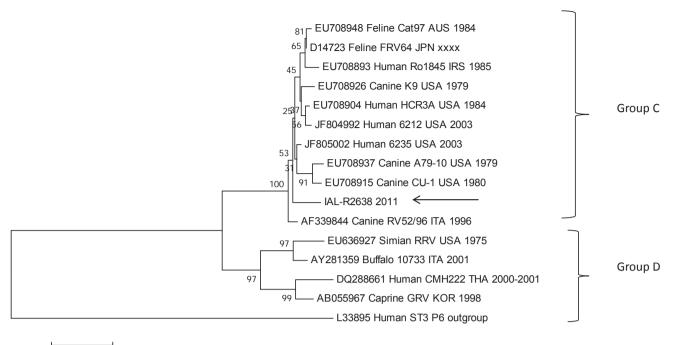


Figure 3 Deduced amino acid sequence of the VP8* trypsin cleavage product of the VP4 protein of human strain IAL-R2638, and of a selection of P[3] rotaviruses. Hyper-variable region (amino acids 71 to 204), the highly conserved cysteine ($\mathbf{\nabla}$), prolines ($\mathbf{\bullet}$), and arginines (\mathbf{n}) are indicated. Species and isolate of each strain are indicated.

Still inside the VP8* variable region, a substitution at residue $155^{Y \rightarrow H}$ was observed in strain IAL-R2638, which was also shared by strains K9 (canine), HCR3A (human) and 6212 (human). Another two amino acid substitutions inside variable region were observed in strain IAL-R2638: $92^{I \rightarrow V}$ and $201^{A \rightarrow T}$ (Figure 3). Other amino acid substitutions were observed outside VP8* hyper-variable region in strain IAL-R2638: (i) an inversion in amino acid residues at positions $233^{V \rightarrow I}$ and $234^{I \rightarrow V}$, and (ii) amino acid substitution at position $245^{S \rightarrow P}$. This last substitution was shared exclusively with human strain Ro1845 (Figure 3).

The overall amino acid sequence identity between the VP4 gene of strain IAL-R2638 and those

cognate P[3] sequences ranged from 84.2 to 96.8%. The VP4 deduced amino acid sequence of IAL-R2638 showed the highest identity to P[3] canine RVA strain RV52/96 (96.8%). Strain IAL-R2638 also exhibited high amino acid identity to feline strain FRV64 (96.4%), to human strain HCR3A (96.4%), and to canine strain K9 (96.4%). The lowest amino acid identity was observed between strain IAL-R2638 and simian strain RRV (84.2%) (Figure 4). In addition, the VP4 deduced amino acid sequence of P[3] strains could be discriminated into two distinct clusters, designated C and D. The Brazilian RVA strain IAL-R2638 clustered into group C, a part of the reference simian strains RRV (EU636927) (Figure 4).



0.05

Figure 4 Neighbor-joining tree of the partial VP4 deduced amino acid sequence generate with MEGA 4.0 software of the IAL-R2638 strain (arrow). Reference P[3] strains were obtained from GenBank database. Accession number, species, isolates, countries and year of each strain are indicated. Group C and D represent two clusters genetically distinct. The scale indicates the number of divergent amino acid residues. Percentages of bootstrap values are shown at the branch node.

DISCUSSION

RVA bearing G3P[3] specificities are common in both cats and dogs.¹⁶ However, there also have been few reports describing the detection of G3P[3] strains in humans,^{7,17,18} including in Brazil.^{8,19,20} The detection of G3P[3] in humans was presumably due to the interspecies transmission from animals to humans,¹⁹ and the analysis of these genomes give general insights into the diversity and evolution of the RVA strains.^{3,7,8,17,22,23}

Previous G3P[3] nucleotide analysis studies have demonstrated the existence of a particular canine-feline genogroup comprising animal (i.e. Cat97, CU-1, K9, A79-10) and animal-derived human (i.e. HCR3A, Ro1845) strains^{16,22,23} that were distinct from genomes of human, simian, bovine or porcine origin.²³ The human G3P[3] strain IAL-R2638 was also shown to belong to this genogroup,⁸ a finding which is confirmed in the present amino acid analysis.

The main hypothesis on the basis of many methods of phylogenetic analysis is that all features undergo evolution independently of each other.¹² However, this supposition could not be completely applied for polypeptides because potentially unfavorable effect of some nucleotide substitutions may be compensated by one or several

substitutions in other positions of the sequence (i.e. coordinated or correlated substitutions).^{24,25} Relatively high proportion of coordinated substitutions may result in significant errors in phylogenetic analysis, and an incorrect topology of a phylogenetic tree.¹² It should be noted that the same topology exhibited by phylogenetic trees in Luchs et al⁸ study was maintained in the present amino acid-based trees, reinforcing the rather stable relationship between G3P[3] strains.

The VP7 hyper variable regions A-F was completely identical between IAL-R2638 and the strains A79-10 (canine), CU-1 (canine), RV198-95 (human), HCR3A (human), Ro1845 (human), and Cat97 (feline); with amino acid sequence identity ranging from 98.8 to 99.2%. Therefore, the amino acid divergence was located outside the variable regions. However, the evolutionary value of this data remains unclear. Interestingly, the amino acid identity of VP7 protein was higher (99.2%) between canine-derived human HCR3A (USA) and IAL-R2638 (Brazil) strains. Taking the risks, it could be suggested that canine RVA might acquire mutations during replication in a heterologous host (i.e. humans). Geographic location may also play a role in this high amino acid identity observed, since both strains were detected in American Continent.

In addition, IAL-R2638 strain shared the amino acid substitutions at positions $147^{A \rightarrow T}$, $212^{A/T \rightarrow V}$, $213^{A/T/N \rightarrow V}$, and $242^{A/N/S \rightarrow V}$ with the canine-feline genogroup strains, including the canine-feline-derived human RVA strains (i.e. HCR3A and Ro1845). The amino acid substitution at position $147^{A \rightarrow T}$ occurred inside of the major antigenic site, region D (amino acids 143 to 152). The amino acid substitution at positions $212^{A/T \rightarrow V}$ and $213^{A/T/N \rightarrow V}$ occurred inside the antigenic site E (amino acid 207 to 220), which is spatially very close to site D. Some authors have been described region E ranging from amino acid 208 to 223/224^{26,27}; therefore, the amino acid substitution at position $221^{A \rightarrow T}$. could also be considered inside antigenic site E. Bearing in mind all data together, those substitutions could modify the antigenicity of the corresponding region, and maybe change the virus interaction with the immune system. The precise impact of amino acid changes cannot be predicted from sequence information alone,²⁸ and studies attempting to correlate intragenotypic nucleotide differences with antigenic differences are extremely important.²⁹

The potential VP7 N-linked glycosylation site located at amino acid 69 in IAL-R2638 strain tends to be conserved among RVA strains.³⁰ Recently, the circulating Belgian G3 RVA strains were found to possibly possess an extra N-linked glycosylation site at amino acid 238.²⁸ Glycosylation of residue 238 could have far-ranging effects on the immunogenicity, and has previously been shown to reduce neutralization of animal RVA strains by hyper immune sera and monoclonal antibodies.²⁶ The G3 RVA vaccine strain of RotaTeq[™] does not display this additional glycosylation site at amino acid 238,²⁸ as well as the G3P[3] canine-feline genogroup strains. Therefore, theoretically, this vaccine could efficiently protect against G3P[3] genotype infections, considering its highly conserved genome origin.

Trypsin cleavage of VP4 spike protein which yields two polypeptides, VP8* and VP5* is required for the activation of infectivity.³¹ The potential VP4 arginine/lysine cleavage sites (231, 241 and 247) were maintained in strain IAL-R2638, ensuring infectivity. The four proline residues (68, 71, 225 and 226) are conserved in IAL-R2638 strain. These conserved prolines may have a major influence on the conformation of VP4, because proline is known to distort three-dimensional structure.³¹ It is known that there is a diversity of VP4 sequences in RVA. Animal RVA strains, including canine and feline P[3] strains, were found to have different amino acids in the positions corresponding to those in common human RVA.³² The previously identified cysteine residue at position 215, proline residues at position 224 and 225, and arginine residues at positions 230, 240, and 246 in human VP4 RVA are found at position 216 (C), 225 (P), 226 (P), 231 (R), 241 (R), and 247 (R), respectively, in animal strains.^{31,32} These differences at amino acids positions were observed in the IAL-R2638 analysis. In fact, a comparison of the VP8* fragment amino acid sequences of seven representative P genotype strains (Wa and WI61 for genotype P[8], DS-1 for genotype P[4], Gottfried and M37 for genotype P[6], K9 and HCR3A for genotype P[3]) indicated a insertion of three nucleotides in P[3] strains: two timines at positions 399 and 400, and one citocine at position 409 (data not shown). Probably, the insertion of these three nucleotides resulted in the addition of an amino acid after position 132.

The present data indicate that the VP4 protein of four P[3] strains (simian RRV, human CMH222, caprine GVR, and buffalo 10733) displayed only 84.2-85% amino acid identities (77.5-77.7% nucleotide identities) with the canine-feline genogroup. Previous phylogenetic analysis of relationship among G3P[3] RVA of VP4 gene have already revealed two major lineages within P[3] genotype, keeping these 4 strains together in a separate group.^{8,18,19,33}

In summary, data from amino acid analysis were consistent with the sequence data of VP7 and VP4 gene segments of the IAL-R2638 strain, supporting its possible canine origin. However, the choice of the variable regions of VP7 and VP4 to serve as evidence for evolutionary relationships between G3P[3] strains was arbitrary; and other combinations would probably give similar results. It is worth mentioning that this study analyzed only two RVA proteins from IAL-R2638 strain, and it is also possible that one or more of its other segments were derived from human RVA strains making it a human-animal reassortant. In addition, the present study raises the question as to whether the amino acid-based trees could be applied as an approach to the study of RVA evolution, avoiding incorrect phylogenetic reconstructions.^{9,10,11,12}

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REFERENCES

- 1. Kapikian AZ, Hoshino Y, Chanock RM. Rotaviruses. In: Knipe DM, Howley RM, Griffin DE, Lamb RA, Martin MA, Roizman B, Straus SE, editors. Fields Virology, 4th ed., Vol 2. Philadelphia: Lippincott Williams and Wilkins; 2001. p. 1787-1833.
- Santos N, Hoshino Y. Global distribution of rotavirus serotypes/ genotypes and its implication for the development and implementation of an effective rotavirus vaccine. Rev Med Virol. 2005 Jan-Feb;15(1):29-56. Review. PubMed PMID: 15484186.
- Wu FT, Bányai K, Lin JS, Wu HS, Hsiung CA, Huang YC, Hwang KP, Jiang B, Gentsch JR. Putative canine origin of rotavirus strain detected in a child with diarrhea, Taiwan. Vector Borne Zoonotic Dis [Internet]. 2012 Feb [cited 2013 Jul 15];12(2):170-3. Available from: http://www.ncbi. nlm.nih.gov/pmc/articles/PMC3267547/pdf/vbz.2011.0708.pdf. doi: http://dx.doi.org/10.1089/vbz.2011.0708. Epub 2011 Oct 24. PubMed PMID: 22022813; PubMed Central PMCID: PMC3267547
- Morillo SG, Luchs A, Cilli A, Costa FF, Carmona Rde C, Timenetsky Mdo C. Characterization of rotavirus strains from day care centers: pre- and post-rotavirus vaccine era. J Pediatr (Rio J) [Internet]. 2010 Mar-Apr [cited 2013 Jul 15];86(2):155-8. Available from: http://www.scielo.br/ pdf/jped/v86n2/en_v86n2a13.pdf. doi: http://dx.doi.org/10.2223/ JPED.1981. PubMed PMID: 20361122.
- Ruiz-Palacios GM, Pérez-Schael I, Velázquez FR, Abate H, Breuer T, Clemens SC, Cheuvart B, Espinoza F, Gillard P, Innis BL, Cervantes Y, Linhares AC, López P, Macías-Parra M, Ortega-Barría E, Richardson V, Rivera-Medina DM, Rivera L, Salinas B, Pavía-Ruz N, Salmerón J, Rüttimann R, Tinoco JC, Rubio P, Nuñez E, Guerrero ML, Yarzábal JP, Damaso S, Tornieporth N, Sáez-Llorens X, Vergara RF, Vesikari T, Bouckenooghe A, Clemens R, De Vos B, O'Ryan M; Human Rotavirus Vaccine Study Group. Safety and efficacy of an attenuated vaccine against severe rotavirus gastroenteritis. N Engl J Med [Internet]. 2006 Jan 5 [cited 2013 Jul 15];354(1):11-22. Available from: http://www.nejm. org/doi/pdf/10.1056/NEJMoa052434. PubMed PMID: 16394298.
- 6. Vesikari T, Matson DO, Dennehy P, Van Damme P, Santosham M, Rodriguez Z, Dallas MJ, Heyse JF, Goveia MG, Black SB, Shinefield HR, Christie CD, Ylitalo S, Itzler RF, Coia ML, Onorato MT, Adeyi BA, Marshall GS, Gothefors L, Campens D, Karvonen A, Watt JP, O'Brien KL, DiNubile MJ, Clark HF, Boslego JW, Offit PA, Heaton PM; Rotavirus Efficacy and Safety Trial (REST) Study Team. Safety and efficacy of a pentavalent human-bovine (WC3) reassortant rotavirus vaccine. N Engl J Med [Internet]. 2006 Jan 5 [cited 2013 Jul 15];354(1):23-33. Available from: http://www.nejm.org/doi/pdf/10.1056/NEJMoa052664. PubMed PMID: 16394299.
- Grant L, Esona M, Gentsch J, Watt J, Reid R, Weatherholtz R, Santosham M, Parashar U, O'Brien K. Detection of G3P[3] and G3P[9] rotavirus strains in American Indian children with evidence of gene reassortment between human and animal rotaviruses. J Med Virol. 2011 Jul;83(7):1288-99. doi: http://dx.doi.org/10.1002/jmv.22076. PubMed PMID: 21567432.
- Luchs A, Cilli A, Morillo SG, Carmona Rde C, Timenetsky Mdo C. Rare G3P[3] rotavirus strain detected in Brazil: possible human-canine interspecies transmission. J Clin Virol. 2012 May;54(1):89-92. doi: http://dx.doi.org/10.1016/j.jcv.2012.01.025. Epub 2012 Mar 6. PubMed PMID: 22398035.
- Matsuda H. Construction of Phylogenetic Trees from Amino Acid Sequences using a Genetic Algorithm. In: Hagiya M, Suyama A, Takagi T, Nakai K, Miyano S, Yokomori T, editors. Proceedings of Genome Informatics Workshop [Internet]; 1995 [cited 2013 Jul 15]; Tokyo (Japan): Universal Academy Press; 1995. p. 19-28. Available from: http://www.jsbi.org/pdfs/journal1/GIW95/Oral/GIW95003.pdf.

- Ahmad S, Selvapandiyan A, Bhatnagar RK. A protein-based phylogenetic tree for gram-positive bacteria derived from hrcA, a unique heat-shock regulatory gene. Int J Syst Bacteriol [Internet]. 1999 Oct [cited 2013 Jul 15];49 Pt 4:1387-94. Available from: http://ijs.sgmjournals.org/ content/49/4/1387.long. PubMed PMID: 10555317.
- Foster PG, Hickey DA. Compositional bias may affect both DNAbased and protein-based phylogenetic reconstructions. J Mol Evol. 1999 Mar;48(3):284-90. doi: http://dx.doi.org/10.1007/PL00006471. PubMed PMID: 10093217.
- Sherbakov DY, Triboy TI. Effect of co-evolving amino acid residues on topology of phylogenetic trees. Biochemistry (Mosc) [Internet]. 2007 Dec [cited 2013 Jul 15];72(12):1363-7. Available from: http://protein. bio.msu.ru/biokhimiya/contents/v72/pdf/bcm_1363.pdf. PubMed PMID: 18205620.
- Gentsch JR, Glass RI, Woods P, Gouvea V, Gorziglia M, Flores J, Das BK, Bhan MK. Identification of group A rotavirus gene 4 types by polymerase chain reaction. J Clin Microbiol [Internet]. 1992 Jun [cited 2013 Jul 15];30(6):1365-73. Available from: http://jcm.asm.org/ content/30/6/1365.long. PubMed PMID: 1320625; PubMed Central PMCID: PMC265294.
- Gouvea V, Glass RI, Woods P, Taniguchi K, Clark HF, Forrester B, Fang ZY. Polymerase chain reaction amplification and typing of rotavirus nucleic acid from stool specimens. J Clin Microbiol [Internet]. 1990 Feb [cited 2013 Jul 15];28(2):276-82. Available from: http://jcm. asm.org/content/28/2/276.long. PubMed PMID: 2155916; PubMed Central PMCID: PMC269590.
- Tamura K, Dudley J, Nei M, Kumar S. MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. Mol Biol Evol [Internet]. 2007 Aug [cited 2013 Jul 15];24(8):1596-9. Available from: http://mbe.oxfordjournals.org/content/24/8/1596.full.pdf+html. Epub 2007 May 7. PubMed PMID: 17488738.
- Martella V, Pratelli A, Elia G, Decaro N, Tempesta M, Buonavoglia C. Isolation and genetic characterization of two G3P5A[3] canine rotavirus strains in Italy. J Virol Methods. 2001 Jul;96(1):43-9. doi: http://dx.doi. org/10.1016/S0166-0934(01)00312-3. PubMed PMID: 11516488.
- De Grazia S, Martella V, Giammanco GM, Gòmara MI, Ramirez S, Cascio A, Colomba C, Arista S. Canine-origin G3P[3] rotavirus strain in child with acute gastroenteritis. Emerg Infect Dis [Internet]. 2007 Jul [cited 2013 Jul 15];13(7):1091-3. Available from: http:// www.ncbi.nlm.nih.gov/pmc/articles/PMC2878246/pdf/07-0239_ finalD.pdf. PubMed PMID: 18214189; PubMed Central PMCID: PMC2878246.
- Khamrin P, Maneekarn N, Peerakome S, Yagyu F, Okitsu S, Ushijima H. Molecular characterization of a rare G3P[3] human rotavirus reassortant strain reveals evidence for multiple human-animal interspecies transmissions. J Med Virol. 2006 Jul;78(7):986-94. doi: http://dx.doi.org/10.1002/jmv.20651. PubMed PMID: 16721863.
- Santos N, Clark HF, Hoshino Y, Gouvea V. Relationship among serotype G3P5A rotavirus strains isolated from different host species. Mol Cell Probes. 1998 Dec;12(6):379-86. doi: http://dx.doi.org/10.1006/ mcpr.1998.0198. PubMed PMID: 9843655.
- Timenetsky Mdo C, Santos N, Gouvea V. Survey of rotavirus G and P types associated with human gastroenteritis in São Paulo, Brazil, from 1986 to 1992. J Clin Microbiol [Internet]. 1994 Oct;32(10):2622-4. Available from: http://jcm.asm.org/ content/32/10/2622.long. PubMed PMID: 7814514; PubMed Central PMCID: PMC264121.

- Nakagomi T, Nakagomi O. Human rotavirus HCR3 possesses a genomic RNA constellation indistinguishable from that of feline and canine rotaviruses. Arch Virol. 2000;145(11):2403-9. doi: http://dx.doi. org/10.1007/s007050070029. PubMed PMID: 11205126.
- 22. Matthijnssens J, De Grazia S, Piessens J, Heylen E, Zeller M, Giammanco GM, Bányai K, Buonavoglia C, Ciarlet M, Martella V, Van Ranst M. Multiple reassortment and interspecies transmission events contribute to the diversity of feline, canine and feline/ canine-like human group A rotavirus strains. Infect Genet Evol. 2011 Aug;11(6):1396-406. doi: http://dx.doi.org/10.1016/j. meegid.2011.05.007. Epub 2011 May 13. PubMed PMID: 21609783.
- 23. Tsugawa T, Hoshino Y. Whole genome sequence and phylogenetic analyses reveal human rotavirus G3P[3] strains Ro1845 and HCR3A are examples of direct virion transmission of canine/feline rotaviruses to humans. Virology. 2008 Oct 25;380(2):344-53. doi: http://dx.doi. org/10.1016/j.virol.2008.07.041. Epub 2008 Sep 11. PubMed PMID: 18789808; PubMed Central PMCID: PMC2575048.
- 24. Hughes AL, Yeager M. Coordinated amino acid changes in the evolution of mammalian defensins. J Mol Evol. 1997 Jun;44(6):675-82. doi: http://dx.doi.org/10.1007/PL00006191. PubMed PMID: 9169560.
- Pazos F, Helmer-Citterich M, Ausiello G, Valencia A. Correlated mutations contain information about protein-protein interaction. J Mol Biol. 1997;271(4):511-23. doi: http://dx.doi.org/10.1006/ jmbi.1997.1198. PubMed PMID:9281423.
- Ciarlet M, Hoshino Y, Liprandi F. Single point mutations may affect the serotype reactivity of serotype G11 porcine rotavirus strains: a widening spectrum? J Virol [Internet]. 1997 Nov [cited 2013 Jul 15];71(11):8213-20. Available from: http://jvi.asm.org/ content/71/11/8213.long. PubMed PMID: 9343172; PubMed Central PMCID: PMC192278.
- Dyall-Smith ML, Lazdins I, Tregear GW, Holmes IH. Location of the major antigenic sites involved in rotavirus serotype-specific neutralization. Proc Natl Acad Sci U S A [Internet]. 1986 May [cited 2013 Jul 15];83(10):3465-8. Available from: http://www.pnas.org/ content/83/10/3465.long. PubMed PMID: 2422651; PubMed Central PMCID: PMC323536.
- 28. Zeller M, Patton JT, Heylen E, De Coster S, Ciarlet M, Van Ranst M, Matthijnssens J. Genetic analyses reveal differences in the VP7 and VP4 antigenic epitopes between human rotaviruses circulating in Belgium and rotaviruses in Rotarix and RotaTeq. J Clin Microbiol

[Internet]. 2012 Mar [cited 2013 Jul 15];50(3):966-76. Available from: http://jcm.asm.org/content/50/3/966.full.pdf+html. doi: http:// dx.doi.org/10.1128/JCM.05590-11. Epub 2011 Dec 21. PubMed PMID: 22189107; PubMed Central PMCID: PMC3295124.

- Hoshino Y, Jones RW, Ross J, Honma S, Santos N, Gentsch JR, Kapikian AZ. Rotavirus serotype G9 strains belonging to VP7 gene phylogenetic sequence lineage 1 may be more suitable for serotype G9 vaccine candidates than those belonging to lineage 2 or 3. J Virol [Internet]. 2004 Jul [cited 2013 Jul 15];78(14):7795-802. Available from: http:// jvi.asm.org/content/78/14/7795.full.pdf+html. PubMed PMID: 15220453; PubMed Central PMCID: PMC434108.
- Martella V, Ciarlet M, Pratelli A, Arista S, Terio V, Elia G, Cavalli A, Gentile M, Decaro N, Greco G, Cafiero MA, Tempesta M, Buonavoglia C. Molecular analysis of the VP7, VP4, VP6, NSP4, and NSP5/6 genes of a buffalo rotavirus strain: identification of the rare P[3] rhesus rotavirus-like VP4 gene allele. J Clin Microbiol [Internet]. 2003 Dec [cited 2013 Jul 15];41(12):5665-75. Available from: http://jcm.asm. org/content/41/12/5665.full.pdf+html. PubMed PMID: 14662959; PubMed Central PMCID: PMC309001.
- Gorziglia M, Green K, Nishikawa K, Taniguchi K, Jones R, Kapikian AZ, Chanock RM. Sequence of the fourth gene of human rotaviruses recovered from asymptomatic or symptomatic infections. J Virol [Internet]. 1988 Aug;62(8):2978-84. Available from: http://jvi.asm. org/content/62/8/2978.long. PubMed PMID: 2839714; PubMed Central PMCID: PMC253736.
- Taniguchi K, Urasawa T, Urasawa S. Species specificity and interspecies relatedness in VP4 genotypes demonstrated by VP4 sequence analysis of equine, feline, and canine rotavirus strains. Virology. 1994 May 1;200(2):390-400. doi: http://dx.doi.org/10.1006/viro.1994.1203. PubMed PMID: 8178429.
- 33. Matthijnssens J, Taraporewala ZF, Yang H, Rao S, Yuan L, Cao D, Hoshino Y, Mertens PP, Carner GR, McNeal M, Sestak K, Van Ranst M, Patton JT. Simian rotaviruses possess divergent gene constellations that originated from interspecies transmission and reassortment. J Virol [Internet]. 2010 Feb [cited 2013 Jul 15];84(4):2013-26. Available from: http://jvi.asm.org/content/84/4/2013.full.pdf+html. doi: http://dx.doi.org.br/10.1128/JVI.02081-09. Epub 2009 Nov 25. PubMed PMID: 19939934; PubMed Central PMCID: PMC2812371.

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